ISOLATION AND STRUCTURES OF A NEW ALKALOID ALATAMINE AND AN INSECTICIDAL ALKALOID WILFORDINE FROM EUONYMUS *ALATUS* FORMA *STRIATUS* (THUNB.) MAKINO.

K. **YAMADA,* Y. SHIZURI** and Y. **HIRATA**

Department of Chemistry, Faculty of Science. Nagoya University, Chikusa, Nagoya, Japan

(Received *in Japan 26 lkcember* **1977;** *Received in the UK for publication 27 February* **1978)**

Abstract-A new alkaloid alatamine 5 and an insecticidal alkaloid wilfordine 6 were isolated together with evonine **2, neocvonine 3, and euonymine 4 from** *Euonymns alafus* **forma striatus (Tbunb.) Makino (family** *Cclasfraceae).* **The structures of alatamine 5 and wilfordine 6 were fully established by chemical and spectroscopic methods. Transformation of alatamine 5 to wilfordine 6 was made.**

Although the presence of alkaloids in the family Celastraceae has for a long time been known,' and isolation^{2,3,9} of a few alkaloids and some structural studies^{3,10} were made, complete structural elucidation of a number of new and previously known alkaloids⁴ and the related new sesquiterpenes' has **been** carried out only in recent years.

As part of our studies on alkaloids' and sesquiterpenoids' of the *Euonymus* genus in the family *Celas*fraceae, we have examined the alkaloidal components of *Euonytnus alatus* forma *striatus* (Thunb.) Makino (Japanese name, Komayumi), and isolated a new alkaloid alatamine 5 and an insecticidal alkaloid wilfordine 6 previously obtained from *Tripegium wilfordii* Hook (family *Celastraceae).* Insecticidal activity of *Tripergium* wilfordii Hook has recorded since 1931,⁸ and efforts to extract the active component(s) were made by Acree and Haller⁹ and by Beroza,^{104,b} resulting in the isolation of wilfordine as one of the insecticidally active alkaloids of this plant. The results of the structural studies of wilfordine by Beroza, which are summarized below, indicate that wilfordine 6 is a C₁₅-decahydroxy compound which is esterified with hydroxywilfordic acid 1, benzoic acid, and 5 moles of acetic acid.^{10a,c} The structure of hydroxywilfordic acid **1** was determined by Beroza.'&

ethereal solution was again dissolved in ether, and the mixture was repeatedly extracted with dil. HCl solution. The aqueous layers were made basic (pH 9) with solid $K₂CO₃$ and extracted with AcOEt. On removal of AcOEt the crude alkaloidal mixture was obtained, which was chromatographed over silicic acid, affording evonine $2^{4b,6a-f}$ neoevonine $3,^{6c-f}$ euonymine $4,^{6c,d,j}$ alatamine 5, and wilfordine 6.¹¹ The physical and spectral properties of the two alkaloids follow:

Alatamine 5: $C_{41}H_{45}NO_{18}$; m.p. 185-193°; $[\alpha]_{D}^{22} + 44^{\circ}$ (c 1.08, CHCl₃); UV, λ_{max} (EtOH), nm (e) 233 (24,400). 272 (5300); IR (CHCb) 3540, 3470, 1750 (broad), 1603, 1588, 1576 cm-'; NMR (Table 1); Mass 839 (molecular ion peak).

Wilfordine 6: C₄₃H₄₉NO₁₉; m.p. 170-176°; $[\alpha]_D^{22} + 5^\circ$ (c 5.0, CHCl₃); UV λ_{max} (EtOH), nm (e) 232 (22,600), 71 (4400); IR (CHCL) 3530, 3460, 1745 (broad), 1602. 1585, 1565 cm⁻¹; NMR (Table 1); Mass 883 (molecular ion peak).

Transformation of alatamine 5 to wiifordine 6, and the structure of the sesquite~ene part of wilfordine 6. Comparison of the molecular formulas and the spectral data of alatamine and wilfordine suggests that a keto **group** in alatamine would exist as a secondary acetoxyl group in wilfordine. This inference was confirmed by

$$
\begin{array}{ccc}\n & & \xrightarrow{+8H_{2}O} & & \xrightarrow{+8H_{
$$

We have determined the structures of alatamine 5 and wilfordine 6, and the results were reported as a short communication.^{on} The present paper describes the details of our structural studies on these alkaloids 5 and 6.

ISOLATION AND STRUCTURES

The fruits of *Euonymus alatus* **forma** *striatus* (Thunb.) Makino were ground in MeOH and the mixture was filtered. The MeOH extract was concentrated to give an aqueous solution, which was shaken with ether repeatedly. The residue obtained on evaporation of the transforming alatamine to wilfordine: reduction of alatamine with NaBH4 in DMF followed by acetylation with Ac20 in pyridine afforded a mixture of two epimers, one of which was identified as wilfordine. Beroza obtained a crystalline C_{15} -compound $(C_{15}H_{26}O_{10})$ by alkaline hydrolysis of wilfordine.¹⁰⁰ We have confirmed the Beroza's results and obtained the C_{15} -compound more **conveniently by reduction of wilfordine with** LAH in THF. This C_{15} -compound was proved to be euonyminol 7, the structure of which was unambiguously established in our laboratory.⁶⁴

Alatamine 5. Considering the Beroza's results on

Table 1. NMR spectral data (δ ppm, 60 MHz, CDCl3)

* This signal appeared in the region of 6 4.5 - 5.5.

wilfordine 6 (vide ante) together with the transformation of alatamine 5 to wilfordine 6, alatamine was shown to be the C_{15} -polyhydroxy compound $(C_{15}H_{24}O_{10})$ which was esterified with hydroxywilfordic acid 1, benzoic acid, and four molec of AcOH. The NMR spectrum of alatamine showed the presence of four acetate groups $[81.95, 2.10, 2.12, 2.24$ (3H each) in CDCl₃], and methanolysis of alatamine (MeONa-MeOH) gave one mole each of methyl benzoate and the dimethyl ester of hydroxywilfordic acid 1, as expected.

1916

In the NMR spectra (Table 1) the signals due to the C₁₅-part of alatamine 5 was shown to correspond well to those of the sesquiterpene part (evoninol) of evonine 2, suggesting the sesquiterpene moiety of alatamine 5 to be evoninol 10. This view was verified by the following findings: (1) alatamine 5 afforded euonyminol octaacetate 8^{6e} and isoeuonyminol octaacetate 9^{6e} in the ratio of 1:2 on reduction with LAH followed by acetylation in the same manner as evonine 2 did;⁶ (2) both alatamine 5 and evonine 2 were transformed to a common evoninol derivative 25 (vide post).

In order to establish the positions of the benzoate and hydroxywilfordate groups in the evoninol nucleus 10 of alatamine 5, experiments included in (A) and (B) were performed.

(A) Concerning the site of the benzoate group in alatamine 5. As shown in (B), hydroxywilfordic acid 1 is attached to C-3 and C-15 of the evoninol nucleus 10 in alatamine, the benzoate group in alatamine was expected to occupy one of the possible five positions (C-1, C-2, C-5, C-8, C-11) in the evoninol nucleus. Thus, three monobenzoates 11, 12, and 13 were synthesized from evonine 2 as model compounds. The monobenzoate 11 was obtained as follows: pentadesacetyl evonine acetonide 14^{6e} was benzoylated with (PhCO)₂O in pyridine to give 15, which was subjected to deacetalization followed by acetylation. The position of the benzoate group in 15 was determined by the downfield shift of the NMR signal due to H-1 from δ 4.24 in 14 to δ 5.90 in 15.

The monobenzoate 12 was synthesized as follows: pentadesacetyl evonine 16^{6e} was acetylated with Ac₂O-AcONa, affording the 2-desacetyl evonine 17, which on benzovlation with PhCOCl in pyridine yielded 12. From the NMR spectrum of 17, OH groups at C-1, C-5, C-8, and C-11 were shown to be acetylated, proving that the position of the benzoate group in 12 is at C-2. The monobenzoate 13 was prepared from pentadesacetyl
evonine triacetate 18:⁶⁴ benzoylation of 18 with (PhCO)₂O in pyridine gave 19, which on acetylation was led to 13. In this case again, the site of benzoylation in 19 was detected by the NMR spectral comparison of 18 and 19.

Concerning the chemical shifts of the NMR signals due to H-1, H-2, H-3, and H-11, and due to four acetate methyls (8 1.95, 2.10, 2.12, 2.24 in alatamine 5; 8 1.52, 2.05, 2.20, 2.26 in 11; 8 1.90, 2.10, 2.10, 2.25 in 12; 8 1.92, 2.06, 2.25, 2.25 in 13), comparison was made among alatamine 5 and these three benzoates (11, 12, and 13),

11: $R' = PhCO, R^2 = R^3 = Ac$ **12:** $R' = R^3 = Ac$, $R^2 = PhCO$ **13:** $R' = R^2 = Ac$, $R^3 = PhCO$

C-2 in alatamine 5. It should be noted that the chemical the δ -lactams 26 [$\nu_{C-O}(CHCl_3)$ 1735, 1640 cm⁻¹], the shifts of four acetate Me signals of alatamine 5 cor-
former being identical in all respects with the spe shifts of four acetate Me signals of alatamine 5 cor-

respond well to those of 12, but not to those of 11 and 13. derived from evonine 2 as described later. respond well to those of 12, but not to those of 11 and 13.

(B) Determination of the sites of the two ester linkages *of hydmxywiljorrfic acid in alatamine 5.* Acetylation of alatamine 5 with Ac_2O -AcONa gave acetyl alatamine 20, which on methanolysis (MeONa-MeOH) yielded 5 desacetyl derivative 21 and 5-desacetyl methyl ester 22. From the NMR spectra of 20 , 21 , and 22 (Table 1) it is evident that deacetylation occurred at C-5 in the reaction, $20 \rightarrow 21 + 22$. The aromatic carboxyl group of hydroxywilfordic acid 1 could be assigned to be attached to C-15 of the evoninol nucleus in alatamine, if one considers in the NMR spectra the chemical shift (8 3.89)"J of the signal **of the** methyl ester in 22 and the upfield shift of the signal arising from H-15 (Table 1) in the reaction $20 \rightarrow 22$.

Acetylation of S-desacetyl methyl ester 22 gave a hexaacetate methyl ester 23. Catalytic hydrogenation of 23 in the presence of $P_tO₂$ in AcOH afforded a piperidine derivative 24, which, on intramolecular aminolysis effected by heating in dioxane, was cleaved into an

suggesting that the benzoate group would be located at evoninol pentaacetate 25 and a diasteromeric mixture of $C-2$ in alatamine 5. It should be noted that the chemical the δ -lactams 26 [$\nu_{C-0}(CHCl_3)$ 1735, 1640 cm⁻

Based on the formation of δ -lactams 26 coupled with the finding that the NMR signal of H-3 underwent the upfield shift in the conversion of 22 *via 23* **and 24 to 25** (Table 1), it is clearly **indicated that** the aliphatic carboxyl group of hydroxywilfordic acid 1 is connected to C-3 of the evoninol nucleus. Therefore, the site of attachment of hydroxywilfordic acid to the evoninol nucleus in alatamine was established. A sequence of reactions $(20 \rightarrow 22 \rightarrow 23 \rightarrow 24 \rightarrow 25 + 26)$ provide a general and unambiguous mean for determining the locations of two ester linkages (macrocyclic bislactone) formed between a sesquiterpene polyol and an unsymmetric dibasic acid, constituents often found in *Celastmceae* alkaloids.

The evoninol derivative 25 discussed above was prepared from evonine 2 as follows. Pentadesacetyl evonine methyl ester 27^{6e} was acelated with Ac₂O-AcONa to give a pentaacetate methyl ester 22, the structure of which was established by the NMR spectral comparison

of 27 and 28: the signal of H-2 was **essentially unchanged, whereas those of H-l, H-S, H-8, H-11, and H-15** shifted downfield in the reaction, $27 \rightarrow 28$. Benzoylation **of 22 with PhCOCl in pyridine afforded a benxoate 29** (Table 1). On catalytic hydrogenation (PtO₂-AcOH) the **benzoate 29 was splif into a stereoisomeric mixture of** the y-lactams 30 $[\nu_{C-O}(\text{CHCl}_3) 1730, 1670 \text{ cm}^{-1}]$ and an **evoninol derivative 25, the latter being found to be** identical with the one 25 obtained from alatamine 5.

Whole structure of alatamine. Formation of the com**mon derivative 26 from alatamine 5 and evonine 2 6rmly established not only the sesquiterpene part of alatamine to be evoninol IO, but also the sites of the benxoate (C-2) and four acetate groupings (C-1, C-5, C-8, C-11) in alatamine. Further, a macrocyclic bislactone in alatamine was proved to be formed between hydroxywilfordic acid** 1 **and two OH groups at C-3 and C-15 of the evoninol nucleus. Consequently the structure of alatamine was determined as 5.**

Wilfordine 6. Considering the transformation of ala**tamine 5 to wilfordine 6 together with the finding that the sesquiterpene part of wilfordine 6 is euonyminol 7 as described above, the structure of wilfordine is represented by 6.**

EXPERIMENTAL

M.ps were uncorrected. UV spectra were measured on a Perkin-Elmer Model 262 spectrophotometer. IR spectra were recorded with JASCO Model IRS and JASCO DS-402G instruments. NMR spectra were obtained using a JNMC-60H instrument: chemical shifts (δ) are reported in ppm downfield from **internal** TMS: signals arising from the sesquiterpene part are cited. Low resolution mass spectra were determined on a Hitachi RMU-6C mass spectrometer equipped with a direct inlet system. High resolution mass spectra were recorded on a JEOLCO GMS-OISG mass spectrometer. Optical rotations were measured on an Oyodenki Model MP-1 spectropolarimeter. For tic and plc silica gel GF₂₅₄, PF₂₅₄ and alumina GF₂₅₄, PF₂₅₄-Type T (E. Merck, A. G., Germany) were used. For column chromatography silicic acid (100 Mesh, Mallinckrodt, U.S.A., and Silica Gel 60, No. 7734, E. Merck, A. G., Germany) were used. Organic solns were washed with sat NaCl aq, dried over Na₂SO₄, and evaporated by vacuum rotary evaporator.

Isolation of alatamine 5 and wilfordine 6. The seeds (ca 28 Kg) of *Euonymus alatus* forma *striatus* (Thunb.) Makino collected in October at Mt. Ibuki (Shigs Prefecture) were ground mechanically in MeOH (151.). The mixture was filtered with suction. The concentrated soln of the filtrate (ca 31.) was extracted with five 3-1 portions of ether. The residue obtained on evaporation of the combined ethereal soln was again dissolved in ether (5 I.). The ethereal soln was extracted with five l-l portions of 2.5% **HCI. The** combined aqueous extracts were made basic

(pH 9) with solid K_2CO_3 , giving ppts. The mixture was extracted with AcOEt five times (5×1) . The combined AcOEt soln was dried and evaporated to afford an oily alkaloidal mixture (9.4g). The mixture was chromatographed on silicic acid (3oog). Tbe column was eluted with solvents of **increasing** polarity. 26Oml fractions being collected [CHCl₃ and MeOH-CHCl₃ (percentages of MeOH: 0.5, I, 2. 3 **and** S%)]. The following alkaloids were obtained, which are listed in the order of elution: evonine 2 $(1.3 g)$, euonymine 4 $(240 mg)$, neoevonine 3 $(60 mg)$, alatamine 5, and wilfordine 6. Alatamine 5 was recrystallized from MeOH to give pure 5 (600 mg). Pure wilfordine 6 (1.2g) was obtained by recrystallization from EtOH. Physical and spectral data of 5 and 6 are **listed in the** text. 5 (Found: C, 58.61; H, 5.28; N. 1.67. C,,H,~NO,s requires: C, 58.64; **H,** 5.40: N, 1.67%). 6 (Found: C, 58.53; H, 5.61; N, 1.52. C₄₃H₄₉NO₁₉ requires: C, 58.44; H, 5.55; N, 1.59%).

Transformation of alatamine 5 *to wilfordinc 6.* **To a soln of 5 (18 mg) in DMF (1 ml) was added NaBH, (4 mg). The** mixture was stirred at 25" for 2.5 hr. added with AcQH (0.05 ml). and extracted with AcOEt. The AcOEt extract was dried and concentrated. A soln of the residue in $Ac₂O$ (0.5 ml) and pyridine (0.5 ml) was stirred at room temp. overnight and concentrated. A soln of the mixture in AcOEt was washed with sat NaHCO₃ aq, dried, and concentrated. The mixture was separated by plc (silica gel) with benzene-AcOEt to give 6 (1.7 mg), m.p. 170-176° after recrystallization from EtOH.

Reduction of wilfonfbte 6 with LAH: Euoaymb~ol 1. A soln of 6 (I5 ma) in THF (2 ml) was added with stirring to a solo of LAH (10 mg) in THF (5 ml) under ice-bath cooling. The mixture was stirred at room temp. for 12 hr and diluted with AcOEt to give ppts. These were filtered off, washed with THF (10 ml), and dissolved in AcOH-H20 (1: 1). The soln was passed through a column of ion-exchange resin Amberlite **IR-120 (H** form). Evaporation of the eluate yielded a colorless residue, recrystallization of which from EtOH afforded pure 7 (6 mg) m.p. 250° (dec). Identification was made by m.m.p. and IR spectral comparison with the authentic specimen.⁶⁰

Reduction of alatamine 5 with LAH: Euonyminol octaacetate 8 and isoeuonyminol octaacetate 9. A soln of 5 (60 mg) in THF (2 ml) **wasaddedtoasolnofLAH(5Omg)inTHF(5mt).Tbemixturcwas stirred at room** temp. for 12 hr and diluted with AcOEt. The ppts were filtered off with suction and washed with THF (10 ml). A soln of the p_{μ} is in AcOH-H₂O (1:1) was passed through a column of ion-exchange resin Amberlite IR-120 (H form), the resin being further eluted with H₂O (100 ml). A solid (25 mg) obtained on evaporation of the eluate was dissolved in $Ac₂O$ (1.5 ml)-pyridine (2 ml), and the soln was stirred at 60' for 12 hr. Evaporation of the mixture gave a residue. which was separated by plc (silica gel) with n-hexane-AcOEt, yielding 12 mg of 8 and 15 mg of 9. Identification of 8 and 9 with authentic specimens was made by m.m.p. and the IR spectral comparison.

Pentadesacetyl evonine acetonide benzoate 15. A soln of 14^{6e}-(98 mg) and (PhCO)₂O (50 mg) in pyridine (2 ml) was kept at 60^o **for 12** hr and concentrated. A residue was **puriiied by plc (silica** gel) with benzene-AcOEt. Recrystallization from MeOH **afforded** I5 (llOmg), m.p. 211-213'; **IR (CHCI,) 3450, 1725** (broad), 1605, 1585, 1575 cm⁻¹; NMR (60 MHz, CDCl₃) 5.90 (1H, d, $J = 3.0$, H-1), 4.13 (1H, dd, $J = 3.0$, 2.8, H-2), 4.87 (1H, d, **J** = 2.8, H-3), 5.25 (1H, d, J = 1.0, H-5), 3.12 (1H, d, J = 1.0, H-6), **4.55 (lH, s,~H-E). 3.76 and 6.01 (2H, ABq. J'= 13.0. H-15);** Mass 695 (M⁺). (Found: C, 62.23; H, 5.95; N, 1.97. C₃₆H₄₁NO₁₃ requires: C, 62.16; H. 5.90: N, 2.01%).

Monobcnzoate **11. A soln of 15 (90 mg) in 5 ml of AcOH-H2O** $(1:1)$ was stirred at 80 $^{\circ}$ for 4 hr under N_2 and concentrated. A **residue was acetylated with** A& **(2 ml)-pyridme (2 ml) at room** temp. for 12 hr. Evaporation of the mixture gave a residue, which was purified by plc (silica gel) with benzene-AcOEt. Recrystallization from EtOH afforded 11 (54 mg), m.p. 183-186°; IR (CHCI~) **3450.1755 (broad). 1605,1585.1575** cm-'; NMR (Table 1): Mass 823 (M⁺). (Found: C, 59.91; H, 5.45; N, 1.79. C₄₁H₄₅NO₁₇ requires: C. 59.78; H. **5.47; N, 1.70%).**

2-*Desacetyl evonine* 17. A mixture of 16^{6e} (200 mg) and AcONa (70 mg) **in A%0 (4 ml) was stirred at 45" for 12 hr, diluted with**

MeOH (3 ml), and concentrated. The residue was separated by plc first by silica gel and subsequently by alumina with benzene-AcOEt, affording 2 (118 mg) and 17 (36 mg; amorphous powder). IR (CHCl₃) 3460, 1755 (broad), 1590, 1575 cm⁻¹; NMR (60 MHz, CDCl₃) 5.57 (1H, d, J = 3.5, H-1), 4.07 (1H, dd, J = 3.5, 3.0, H-2), 4.81 (1H, d, $J = 3.0$, H-3), 6.75 (1H, d, $J = 1.0$, H-5), 3.05 (1H, d, J= 1.0, H-6), 5.57 (HI, s, H-8); 1.65 (3H. d, J= 1.0, H-12). 1.57 $(3H, s, H-14), 3.75$ and 6.03 (2H, ABq, J = 11.0, H-15); Mass 719. (Found: C, 56.90; H, 5.68; N, 1.99. C₃₄H₄₁NO₁₆ requires: C, 56.74 H, 5.74; N, 1.95%).

 $Monobenzoate$ 12. A soln of 17 (36 mg) and PhCOCI (0.05 ml) in pyridine (1.5 ml) was stirred at 45° for 36 hr and concentrated. A CHCl₃ soln of the residue was washed with sat NaHCO₃ aq, dried and concentrated. The residual mixture was separated twice by plc (silica gel) with benzene-AcOEt to give 17 (18 mg) and 12 (15 mg), the latter being recrystallized from MeOH. 12 (11 mg), m.p. 170-1780, IR (CHCls) 3450, 1750 (broad), 1605, 1585, 1575 cm^{-1} ; NMR (Table 1); Mass 823 (M⁻). (Found: C, 60.11; H, 5.58; N, 1.63. C₄₁H₄₅NO₁₇ requires: C, 59.78; H, 5.47; N, 1.70%).

Pentadesacetyl evonine triacetate benzoate 19. A soln of 18⁶⁴ (50 mg) and (PhCO)₂O (20 mg) in pyridine (1 ml) was kept at 50 $^{\circ}$ for 12 hr and concentrated. A CHCl₃ soln of the residue was washed with sat NaHCO₃ aq, dried, and concentrated. The residue was purified by plc (silica gel) with benzene-AcOEt. affording 19 (46 mg, amorphous powder). IR (CHCl₃) 3440, 1755, 1735, 1605, 1590, 1575 cm⁻¹; NMR (60 MHz, CDCl₃) 5.92 (1H, d, $J = 3.5$, H-1), 5.25 (1H, dd, J = 3.5, 3.0, H-2), 4.85 (1H, d, J = 3.0, H-3), 6.68 (1H, d, $J = 1.0$, H-5), 3.26 (1H, d, $J = 1.0$, H-6), 4.61 (1H, s, H-8), 4.66 and 4.97 (2H, ABq, J = 12.5, H-11), 1.65 (3H, s, H-12). 1.56 (3H. s, H-14), 3.75 and 6.04 (2H, ABq, J = 11.0, H-15); Mass 781 (M⁺). (Found: C, 60.23; H, 5.78; N, 1.54. C₃₉H₄₃NO₁₆ requires: C, 59.92; H, 5.51; N, 1.66%).

Monobenzoate 13. A soln of 19 (40 mg) in Ac₂O (2 ml)-pyridine (2 ml) was kept at room temp. for 12hr and concentrated. Recrystallixation of the residue from EtOH gave 13 (30 mg). m.p. 176-184°; IR (CHCl₃) 3520, 1760, 1730, 1605, 1590, 1580 cm⁻ NMR (Table 1); Mass 823 (M⁺). (Found: C, 59.73; H, 5.62; N, 1.64. $C_{41}H_{45}NO_{17}$ requires: C, 59.78; H, 5.47; N, 1.70%).

Acetyl alatamine 20. A mixture of 5 (120 mg) and AcONa (50 mg) in Ac₂O (2.5 ml) was stirred at 60° for 18 hr and concentrated. A CHCI, soln of the mixture was washed with sat NaHCO₃ aq, dried, and concentrated. Recrystallization of the residue from EtOH gave 20 (118 mg), m.p. 183-189°; IR (CHCl₃) 3450, 1745 (broad), 1600, 1585, 1573 cm⁻¹; NMR (Table 1); Mass 881 (M⁺). (Found: C, 58.15; H, 5.22; N, 1.52. C₄₃H₄₇NO₁₉ requires: C, 58.47; H, 5.37; N, 1.59%).

Methanolysis of acetyl alatamine 20: 5-Desacetyl derivative 21 and 5-desacetyl methyl ester 22. To a soln of 20 (100 mg) in MeOH (8 ml) was added a soln (0.05 ml) of 0.75% NaOMe in MeOH under N_2 . The soln was stirred at 0° for 30 min, added to AcOH (0.1 ml), and concentrated. The residual mixture was separated by plc (silica ge1) with benzene-AcGEt, giving crystalline 21(40 mg) and amorphous 22 (44 mg). Recrystallization of the former from MeOH-H₂O afforded pure 21 (35 mg), m.p. 179-184°; IR (CHCl₃) 3380, 1760, 1738, 1598, 1585, 1573 cm⁻¹; NMR (Table 1); Mass 839 (M⁺). (Found: C, 58.78; H, 5.30; N, 1.73. C₄₁H₄₅NO₁₈ requires: C, 58.64; H, 5.40; N, 1.67%). 22; IR (CHCl₃) 3450, 1760, 1740, 1600, 1588, 1575 cm^{-1} ; NMR (Table 1); Mass 871 (M⁺). [High resolution mass spectrum. Found: 871.2917 (M⁺). C₄₂H₄₉NO₁₉ requires: 871.2898].

Hexaacetate methyl ester 23. A soln of 22 (30 mg) in $Ac₂O$ (1.5 ml) and pyridine (1.5 ml) was stirred at room temp. for 12hr and concentrated. Purification of the product by plc (silica gel) with benzene-AcOEt gave amorphous 23 (33 mg); IR (CHCl₃) 3550. 1750 (broad), 1600, 1588, 157Scm"': Mass 955 (M*). [High resolution mass spectrum. Found: 955.3136 (M⁺). $C_{46}H_{53}NO_{21}$ requires: 955.31101.

Formation of evoninol pentaacetate 25 and 8-lactams 26. A soln of 23 (22 mg) in AcOH (3 ml) in the presence of PtO₂ (4 mg) was stirred in the atmosphere of H_2 at room temp. for 12 hr. After removal of the catalyst by filtration, the filtrate was concentrated, giving a piperidine derivative 24, which was dissolved in dioxane (4 ml) and heated at 100° for 12 hr. On removal of solvent there remained a resinous mixture, which was chromatographed on silicic acid with benzene-AcOEt, affording amorphous 25 (10 mg) and δ -lactams 26 (6 mg). 25: IR (CHCl₃) 3520, 1750 cm⁻¹; NMR (Table 1); Mass 611 [M⁺ - 73 *(CH₂OAc)*]. [High resolution mass spectrum. Found: $611.2359 (M^+ - 73)$. $C_{29}H_{39}O_{14}$ requires: 611.2339]. 26 (four stereoisomers, two of which being the major ones according to the NMR **spectral** analysis): IR (CHCl₃) 1735, 1640 cm⁻¹; Mass 283 (M⁺). [High resolution mass spectrum. Found: 283.1401 (M^+). C₁₄H₂₁NO₅ requires: 283.1419].

Pentaacetate methyl ester 28. A mixture of 27^{6s} (300 mg) and AcONa (100 mg) in Ac₂O (10 ml) was stirred at 45° for 12 hr. diluted with MeOH (5 ml) , and concentrated. The residue was purified twice by plc (silica gel and alumina) with benzene-AcOEt, affording the starting 27 (186 mg) and amorphous 28 (48 mg); IR (CHCl₃) 3460, 1750 (Broad), 1590, 1575 cm⁻¹; NMR (Table 1); Mass 793 (M⁺).

Benzoate 29. A soln of 28 (40 mg) and PhCOCl (0.05 ml) in pyridine (2 ml) was stirred at 45° for 48 hr and concentrated. A $CHCl₃$ soln of the residue was washed with sat NaHCO₃ aq, dried, and concentrated. The product was purified by plc (silica gel) twice with benzene-AcOEt, affording amorphous 29 (18 mg); IR (CHCl₃) 3400, 1750, 1603, 1590, 1575 cm⁻¹; NMR (Table 1); Mass 897 (M⁺). [High resolution mass spectrum. Found: 897.3041 (M⁺). C₄₄H₅₁NO₁₉ requires: 897.3055].

Formation of evoninol pentaacetate 25 and y-lactams 39. A sola of 29 (15 mg) in AcOH (2.5 ml) in the presence of P_1O_2 (4 mg) was stirred in the atmosphere of $H₂$ at room temp. for 12 hr. The catalyst was removed by filtration, and the filtrate was concentrated. The mixture was separated by plc (silica gel) with benzene-AcOEt, affording 25 (6 mg) and γ -lactams 30 (3.5 mg). 30 (three stereoisomers **according** to the NMR spectral analysis); IR (CHCis) 1738, 1675 cm $^{-1}$; Mass 225 (M⁺). [High resolution mass spectrum. Found: 225.1345 (M⁺). C₁₂H₁₉NO₃ requires: 225.1364].

Acknowledgements-Financial support for this project from the Ministry of Education, Science, and Culture (Japan), the National Institutes of Health (Grant No. 5 ROl GMO7%9. U.S.A.), and the Kurata Foundation is gratefully acknowledged:

REFERENCES

¹A. Orechoff, Arch. Pharm. 272, 673 (1934).

²K. Doebel and T. Reichstein, *Helv. Chim. Acta* 32, 592 (1949).

- ^{3a}M. Pailer and R. Libiseller. Monatsh. Chem. 93, 403 (1962); b Ibid. 93, 511 (1962); c R. Libiseller and A. Preisinger, Ibid. 93, 417 (1962).
- ⁴⁴ S. M. Kupchan, R. M. Smith and R. F. Bryan, J. Am. Chem. Soc. 92, 6667 (1970); ^bM. Pailer, W. Streicher and J. Leitich, Monatsh. Chem. 102, 1873 (1971); 'A. Klásek, F. Šantavý, A. M. Duffield and T. Reichstein, *Helv. Chim. Acta 54*, 2144 (1971); "A. Klásck, Z. Samek and F. Santavý, *Tetrahed* Letters 941 (1972); 'H. Budzikiewicz, A. Römer and K. Taraz, Z *Natwfotvch. 27b, 800* (1972); 'L. Crombie, P. J. Ham and D. A. Whiting, Phytochem. 12, 703 (1973); ^sL. Dúbravková, L. Dolejš and J. Tomko, Coll. Czechoslov. Chem. Comm. 38, 2132 (1973); ^{*}L. Dúbravková, J. Tomko and L. Dolejš, Phytochem. 12.944 (1973); 'H. Luftmann and G. Spiteller, *Tetrahedmn 30,* 2577 (1974); ^{*i*}H. Wagner, E. Heckel and J. Sonnenbichler, *Tdmhedron Zkttere* 213 (1974); *'Idem,, Tetrahedmn 31.* 1949 (1975); [']M. Cais, D. Ginsberg, A. Mandelbaum and R. M. Smith, Ibid. 31, 2727 (1975); "R. L. Baxter, L. Crombie, D. J. Simmonds and D. A. whiting. 1. *Chem. Sot. Chem. Comm.* 463 *W76k* "Ibid. 465 (1976); "H. Wagner. R. Brgning. H. Latter and A. Jones, *Tetrahedron Letters* 125 (1977): ^PS. M. Kupchan and R. M. Smith, *J. Org. Chem.* **42**, 115 (1977).

^{5a}H. J. den Hertog, Jr., J. Th. Hackmann, D. D. Nanavati and S. Dev, Tetrahedron Letters 845 (1973); ⁶H. J. den Hertog, Jr., C. Kruk. D. D. Nanavati and S. Dev, *Ibid. 2219 (1974); 'H.* Budzikiewicz and A. Römer, *Tetrahedron* 31, 1761 (1975): ⁴C. R. Smith, Jr., R. W. Miller, D. Weisleder, W. K. Rohwedder, N. Eickman and J. Clardy, i. Org. *Chem.* 41, 3264 (1976); 'A. Izbhmer, H. Thomas and H. Budxikiewicz, 2. *Naturfomch.* 31b. 607 (1976).

^{6ª}H. Wada, Y. Shizuri, K. Yamada and Y. Hirata, Tetrahedron Letters 2665 (1971); ^bY. Shizuri, H. Wada, K. Sugiura, K. Yamada and Y. Hirata, Ibid. 2659 (1971); 'K. Sugiura, Y. Shizuri, H. Wada, K. Yamada and Y. Hirata, Ibid. 2733 (1971);
⁴H. Wada, Y. Shizuri, K. Sugiura, K. Yamada and Y. Hirata, Ibid. 3131 (1971); 'Y. Shizuri, H. Wada, K. Sugiura, K. Yamada and Y. Hirata. Tetrahedron 29. 1773 (1973); ¹Y. Shizuri, H. Wada, K. Yamada and Y. Hirata, Ibid. 29, 1795 (1973); ^{*}K. Sugiura, K. Yamada and Y. Hirata, Tetrahedron Letters 113 (1973); ^kY. Shizuri, K. Yamada and Y. Hirata, *Ibid.* 741 (1973); K. Sugiura, K. Yamada and Y. Hirata, Chemistry Letters 579 (1975); ^IK. Yamada, K. Sugiura, Y. Shizuri, H. Wada and Y. Hirata, Tetrahedron 33, 1725 (1977).

⁷⁴ K. Sugiura, Y. Shizuri, K. Yamada and Y. Hirata, Chemistry

Letters 471 (1975); ^bK. Sugiura, Y. Shizuri, K. Yamada and Y. Hirata, Tetrahedron Letters 2307 (1975).

- ⁸ History: W. T. Swingle, H. L. Haller, E. H. Siegler and M. C. Swingle, Science 93, 60 (1941); and references therein.
- ⁹F. Acree, Jr. and H. L. Haller, J. Am. Chem. Soc. 72, 1608 $(1950).$

^{10a} M. Beroza, J. Am. Chem. Soc. 73, 3656 (1951); ^aIbid. 74, 1585 (1952); 'Ibid. 75, 44 (1953); 'Ibid. 75, 2136 (1953); 'J. Org. Chem. 28, 3562 (1963).

¹¹Identity of our alkaloid with wilfordine from Tripergium wilfordii Hook was proved by the mixture melting point and the spectral (IR and mass) comparison. We are grateful to Dr. M. Beroza (USDA, Beltsville, Maryland, U.S.A.) for supplying the generous sample of wilfordine.